

Stage-specific changes in fetal thymocyte proliferation during the CD4⁻8⁻ to CD4⁺8⁺ transition in wild type, *Rag1*^{-/-}, and *Hoxa3*,*Pax1* mutant mice

Dong-ming Su and Nancy R Manley*

Address: Department of Genetics, University of Georgia, Athens, Georgia 30602 USA

E-mail: Dong-ming Su - dmsu@arches.uga.edu; Nancy R Manley* - nmanley@arches.uga.edu

*Corresponding author

Published: 19 September 2002

BMC Immunology 2002, 3:12

Received: 16 July 2002

Accepted: 19 September 2002

This article is available from: <http://www.biomedcentral.com/1471-2172/3/12>

© 2002 Su and Manley; licensee BioMed Central Ltd. This article is published in Open Access: verbatim copying and redistribution of this article are permitted in all media for any purpose, provided this notice is preserved along with the article's original URL.

Abstract

Background: The function of the thymic microenvironment is to promote thymocyte maturation, in part via regulation of thymocyte proliferation and cell death. Defects in fetal thymic epithelial cell (TEC) development and function, and therefore in the formation of a functional microenvironment, can be caused either directly by TEC differentiation defects or indirectly by defective thymocyte maturation. In this paper we studied fetal thymocyte proliferation during the early transition from the CD3⁻4⁻8⁻ (triple negative, TN) to CD4⁺8⁺ (double positive, DP) stages. We compared wild type mice with *Rag1*^{-/-} mice and with *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutant mice, which have blocks at different stages of thymocyte development.

Results: Wild type fetal and adult thymus showed stage-specific differences in the proliferation profiles of developing thymocytes, with fetal stages showing generally higher levels of proliferation. The proliferation profile of fetal thymocytes from *Rag1*^{-/-} mutants also had stage-specific increases in proliferation compared to wild type fetal thymocytes, in contrast to the lower proliferation previously reported for thymocytes from adult *Rag1*^{-/-} mutants. We have previously shown that *Hoxa3*^{+/-}*Pax1*^{-/-} mice have abnormal fetal TEC development, resulting in increased apoptosis at the TN to DP transition and decreased DP cell numbers. Fetal thymocytes from *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutants had increased proliferation, but fewer proliferating cells, at the DP stage. We also observed a decrease in the level of the cytokines IL-7 and SCF produced by *Hoxa3*^{+/-}*Pax1*^{-/-} TECs.

Conclusion: Our results indicate complex and stage-specific effects of abnormal TEC development on thymocyte proliferation.

Background

Thymocyte development occurs within a complex net-

work of cells, extracellular matrix, and secreted factors referred to as the thymic microenvironment [1]. Current

evidence suggests that this microenvironment is created during fetal thymus organogenesis through stage-dependent interactions between differentiating thymic epithelial cells (TECs) and developing thymocytes [2–6]. Therefore, blocks at different stages of thymocyte development result in differential effects on TEC differentiation and function. Because of the complex interdependence of TEC and thymocyte differentiation, defects in fetal TEC development can be caused either directly by cell-autonomous TEC differentiation defects, or indirectly by defective thymocyte maturation. Once a mature thymic environment is formed, many, if not all stages of thymocyte maturation require interactions with thymic epithelial cells. Therefore, defects in TECs can in turn cause defects in thymocyte differentiation, proliferation, or cell death.

Thymocyte differentiation undergoes two main transitions. The first is from the CD3⁻4⁻8⁻ (triple negative, TN) to the CD4⁺8⁺ (double positive, DP) stages. During this transition, T cell precursors sequentially gain and lose cell surface expression of CD44 and CD25 markers, express the pre-T cell receptor (pTCR), and undergo β -selection [2,7–9]. The other transition is from CD4⁺8⁺ DP to CD4⁺ or CD8⁺ single positive (SP) thymocytes, in which DP cells express CD3, rearrange their α -TCR, undergo positive and negative selection, and down regulate CD4 or CD8 to become functional CD8⁺ or CD4⁺ SP T cells [10,11]. In addition to functional and cell surface marker changes, each thymocyte differentiation stage is marked by characteristic changes in proliferation and apoptosis [12–18].

The most well defined role for TECs in directly affecting thymocyte differentiation is to mediate positive selection via expression of MHC molecules [11,19–22]. However, it is still unclear whether TECs directly promote specific stages of thymocyte differentiation during stages that do not involve MHC-TCR interactions, or whether their role at other stages is primarily to promote cell growth and/or survival. Cytokines produced by TECs have been shown to promote proliferation in early stages of thymocyte development, and have been proposed to regulate apoptosis and differentiation, although their precise roles remain controversial [23,24]. TECs have also been shown to promote thymocyte proliferation in SP cells after selection has occurred [15]. However, a defined role for TECs at the TN-DP transition remains elusive. Much of the research on this transition has focused on the role of the pre-TCR. Mounting evidence suggests that the pre-TCR does not require interaction with a ligand to perform its function, and the subsequent CD44⁺25⁻ (TN4 or pre-DP) to DP transition has been termed "programmed differentiation" to indicate that this step can occur in isolated thymocytes, presumably without input from extrinsic signals [25–27]. However, there is also evidence that the thymic microenvironment does contribute signals required for the TN-DP

transition, particularly to promote thymocyte proliferation and apoptosis [13,28–30]. The specific nature and purpose of the TEC contribution during this important transition therefore remains to be determined.

In order to investigate the regulation of thymocyte development by TECs, we are studying *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutant mice. *Hoxa3* and *Pax1* are transcription factors that act in a common pathway regulating thymus organogenesis and thymic epithelial cell development [29,31–34]. *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutants have defects in TEC development including reduced numbers of TECs, fewer MHC Class II⁺ TECs, and changes in TEC proliferation and apoptosis. These TEC defects result in thymocyte maturation defects that include a decrease in total thymocyte number and a 10-fold reduction in the number of DP thymocytes, but an apparently normal transition from the TN3 to the pre-DP stage. The decrease in DP cells is associated with increased apoptosis in both pre-DP and DP cells [29]. As the defects at the TN to DP transition in these mutants are caused by defective TEC function rather than thymocyte-intrinsic defects, these mice represent an opportunity to study the role of TECs in promoting this transition.

To further investigate the role of TECs in promoting thymocyte death and growth during the TN to DP transition, we assayed the proliferation of specific thymocyte subsets using BrdU incorporation, which detects cycling cells in S-phase. We compared fetal and adult wild type mice with *Hoxa3*,*Pax1* compound mutants [29,34]. We also assayed *Rag1*^{-/-} mice, which have defects in TEC development due to arrested thymocyte maturation at the TN3 stage [35]. We found stage-specific differences in proliferation levels between fetal and adult wild type thymocytes. We also found distinct patterns of thymocyte proliferation in the *Rag1*^{-/-} and *Hoxa3*,*Pax1* mutants. The abnormal cell death and proliferation patterns in the *Hoxa3*,*Pax1* compound mutant fetal thymocytes might be caused in part by observed reductions in the transcription levels of the cytokines IL-7 and SCF produced by TECs. Our results support a critical role for microenvironment created by TECs in regulating thymocyte proliferation during the TN to DP transition, and show that this function is disrupted in *Hoxa3*,*Pax1* compound mutants.

Results

Thymocyte proliferation patterns are different in fetal and adult wild type mice

As an initial analysis, we compared thymocyte proliferation between thymocytes from late fetal (E17.5) and adult (6–8 week) wild type C57BL6/J mice. Although differences in the proliferation of fetal vs. adult TCR-expressing thymocytes and have been reported [36], we were interested in investigating the more immature thymocyte subsets.

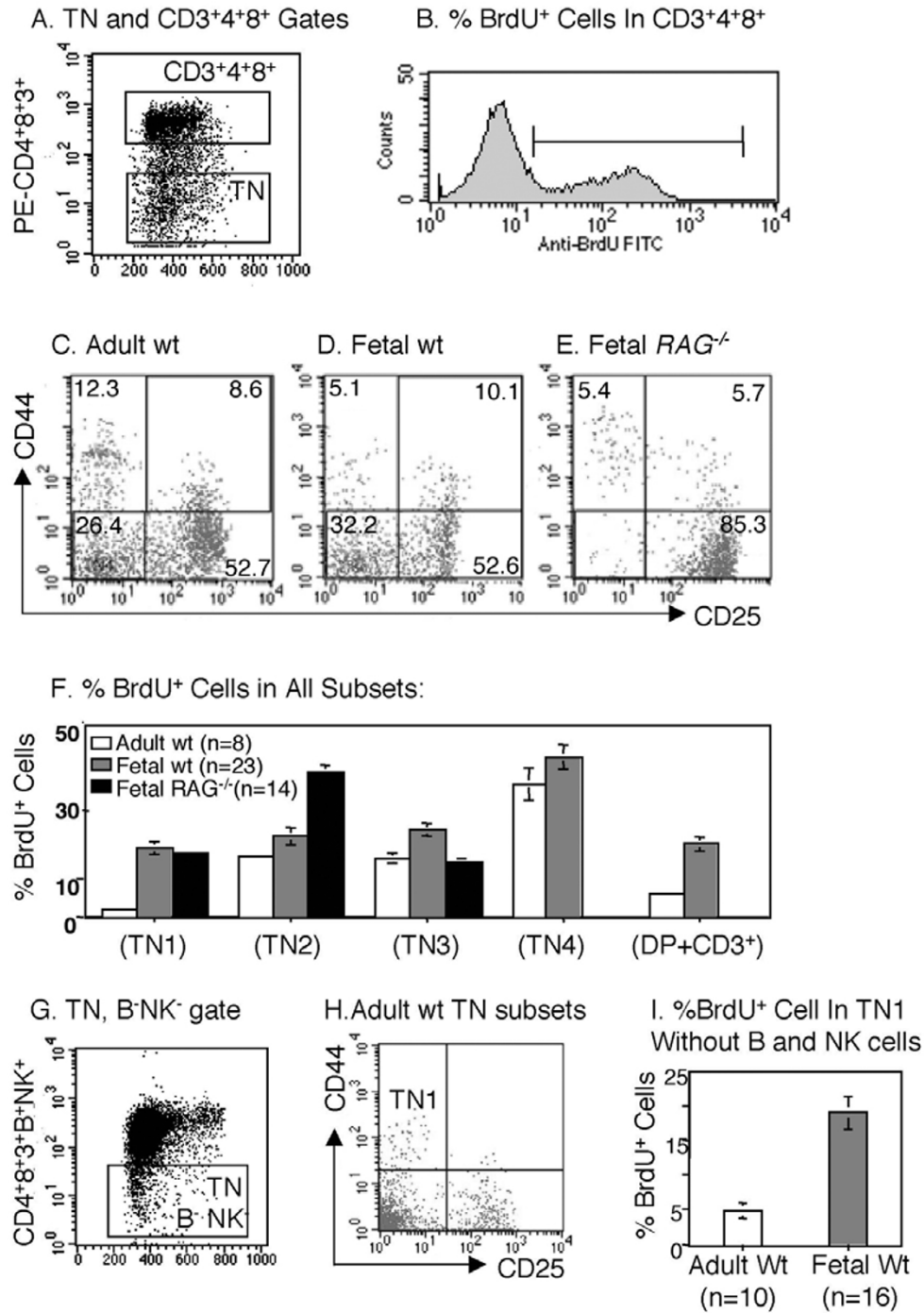


Figure 1
DNA synthesis patterns of fetal vs. adult thymocyte subsets as measured by incorporation of BrdU A. Thymocytes were gated as either CD3⁺4⁺8⁻ TN or CD3⁺4⁺8⁺ DP by cell surface marker expression and forward scatter (SP cells should be located between these two gates, and therefore excluded from analysis). B. Gate used for analysis of BrdU incorporation on gated subpopulations. Panels C-E show CD44 vs. CD25 dot plots of the gated CD3⁺4⁺8⁻ population from 5–6 week old adult C57BL/6 (C), E17.5 fetal C57BL/6 (D), and E17.5 fetal RAG^{-/-} (E). Percentages are shown in each quadrant. F. Percentages of BrdU⁺ cells (Mean ± SEM) in each subset of TN cells defined by CD44 and CD25, and in DP cells. G. Adult wild-type thymocytes were gated as CD3⁺4⁺8⁻B220⁻NK1.1⁻. H. CD44 vs. CD25 dot plots of the gated population from (G). I. Percentage of BrdU⁺ cells (mean ± SEM) in adult and fetal TN1 thymocytes excluding B and NK cells (H, G).

Table 1: Percentage (Mean ± SD) of BrdU⁺ cells in each subset of fetal thymocytes

Group	Total Cell #(X10 ⁵)	No.	CD44 ⁺ CD25 ⁻	CD44 ⁺ CD25 ⁺	CD44 ⁻ CD25 ⁺	CD44 ⁻ CD25 ⁻	CD4 ⁺ CD8 ⁺ CD3 ⁺	In All DN
wt Adult	NC	8	2.04 ± 0.53*	15.83 ± 1.73*	15.38 ± 3.36*	34.73 ± 11.43	6.07 ± 1.19*	18.4 ± 5.14*
wt Fetal	42.72 ± 3.88	23	18.05 ± 8.67	20.99 ± 10.66	22.82 ± 8.33	41.78 ± 14.87	19.05 ± 8.12	27.6 ± 8.79
<i>Pax1</i> ^{+/-}	32.14 ± 2.15	22	14.86 ± 8.97	19.12 ± 6.985	16.70 ± 5.47	35.32 ± 14.82	20.80 ± 10.0	24.3 ± 7.38
<i>Hoxa3</i> ^{+/-}	29.25 ± 3.43	13	18.06 ± 9.20	22.87 ± 11.45	19.14 ± 8.73	29.89 ± 15.17	17.02 ± 10.04	21.2 ± 7.23
<i>Pax1</i> ^{-/-}	22.38 ± 4.98	4	18.97 ± 11.50	21.67 ± 4.10	26.31 ± 7.35	45.59 ± 16.90	28.61 ± 9.46	24.8 ± 9.05
<i>Hoxa3</i> ^{+/-} <i>Pax1</i> ^{-/-}	7.96 ± 1.12	6	19.58 ± 8.52	25.28 ± 5.55	22.79 ± 4.56	44.87 ± 14.15	35.37 ± 6.41 ⁴	25.8 ± 6.54
RAG ^{null}	UC	14	16.57 ± 3.12	37.70 ± 7.89**	14.33 ± 3.16**	-	-	11.2 ± 3.3**

* P < 0.01, significant differences between wild type fetal and adult.

** P < 0.01, significant differences with fetal wild type.

*** NC: not count.

BrdU labeled fetal and adult thymi were analyzed for the percent of proliferating cells in different CD3⁺4⁺8⁺ TN thymocyte subsets defined by CD44 and CD25 expression [8]. The CD3⁺4⁺8⁺ population was defined by co-staining of all three markers (Fig. 1A). A typical analysis of the BrdU⁺ cells in DP cells and the four CD44 vs. CD25 subsets within the TN cell population is shown in Figure 1. Compiled data for each subset from all experiments is shown in Table 1.

The wild type adult (Fig. 1C, 1F) and fetal (Fig. 1D, 1F) thymocyte proliferation profile patterns differed significantly at specific stages. The overall proliferation level in TN cells was increased by 50% in fetal thymocytes (p < 0.01). Most dramatically, the percentages of BrdU⁺ cells in fetal thymocytes were 10-fold higher than in the adult at the earliest, TN1 (CD44⁺25⁻) thymocyte differentiation stage (P < 0.001). A much lower, but still significant increase was seen in fetal TN2 cells (25% increase, p < 0.01). Proliferation was also significantly higher at the fetal TN3 stage compared to adult (50% increase, p < 0.01). Proliferation at the TN4/pre-DP (CD44⁻25⁻) stage was similar between fetal and adult thymocytes. In contrast, DP cells showed a 3-fold increase in proliferation in the fetal thymus relative to adult (p < 0.01). Therefore, nearly every stage from the earliest TN to the DP stages showed significantly increased proliferation in fetal thymocytes relative to adults.

It was possible that the lower percentage of proliferating cells seen in the TN1 population in adults was due to an increased number of B cells and NK cells in the adult thymus, which would not have been excluded in our initial analysis. As this was one of the most striking differences seen between fetal and adult stages, we repeated this analysis adding antibodies against B220 and NK1.1 to CD3, 4 and 8 in the excluded gate (Fig. 1G, 1H). Even excluding these cell populations, there is still a significantly lower

percentage of proliferating cells in the TN1 population in the adult thymus than in the fetal thymus (Fig. 1I). Therefore, this difference is not due to different numbers of B and NK cells in the adult vs. fetal thymus.

***Rag1*^{-/-} fetal thymocytes have a stage-specific increase in proliferation relative to wild type**

We also compared proliferation of fetal thymocytes from *Rag1*^{-/-} mice to the results obtained from wild type mice. Thymocytes in the adult *Rag1*^{-/-} thymus have been previously shown to have overall lower levels of proliferation than adult wild type TN thymocytes [13], particularly at the TN3 stage [37–40]. At fetal stages, our results showed that proliferation also varied between thymocytes in the wild type and *Rag1*^{-/-} thymus at discrete stages of thymocyte differentiation. At the earliest TN1 stage, thymocyte proliferation levels in *Rag1*^{-/-} thymi were similar to fetal wild type cells (Fig. 1E, 1F; Table 1). At the TN2 stage, fetal thymocytes from *Rag1*^{-/-} mice had a proliferation level nearly twice that of wild type fetal thymocytes (p < 0.001). By the TN3 stage, the percentage of proliferating *Rag1*^{-/-} thymocytes had dropped slightly below that of wild type fetal thymocytes (~30% decrease, p < 0.01), equal to wild type adult TN3 proliferation levels. In addition, the percentages of BrdU⁺ cells in TN1 and TN2 subsets in the *Rag1*^{-/-} fetal thymus were significantly higher than those of the wild type adults (P < 0.001). These results with *Rag1*^{-/-} fetal thymocytes contrast with thymocytes in the *Rag1*^{-/-} adult, which have much lower proliferation than wild type adult thymocytes at all stages of thymocyte differentiation [13].

Thymocyte proliferation in *Hoxa3*^{+/-}*Pax1*^{-/-} thymocytes at the TN-DP transition

Our previous analysis of the *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutant thymocyte phenotype showed a specific partial block at the TN to DP transition, associated with increased apoptosis at both the TN4/pre-DP and DP subsets, and a

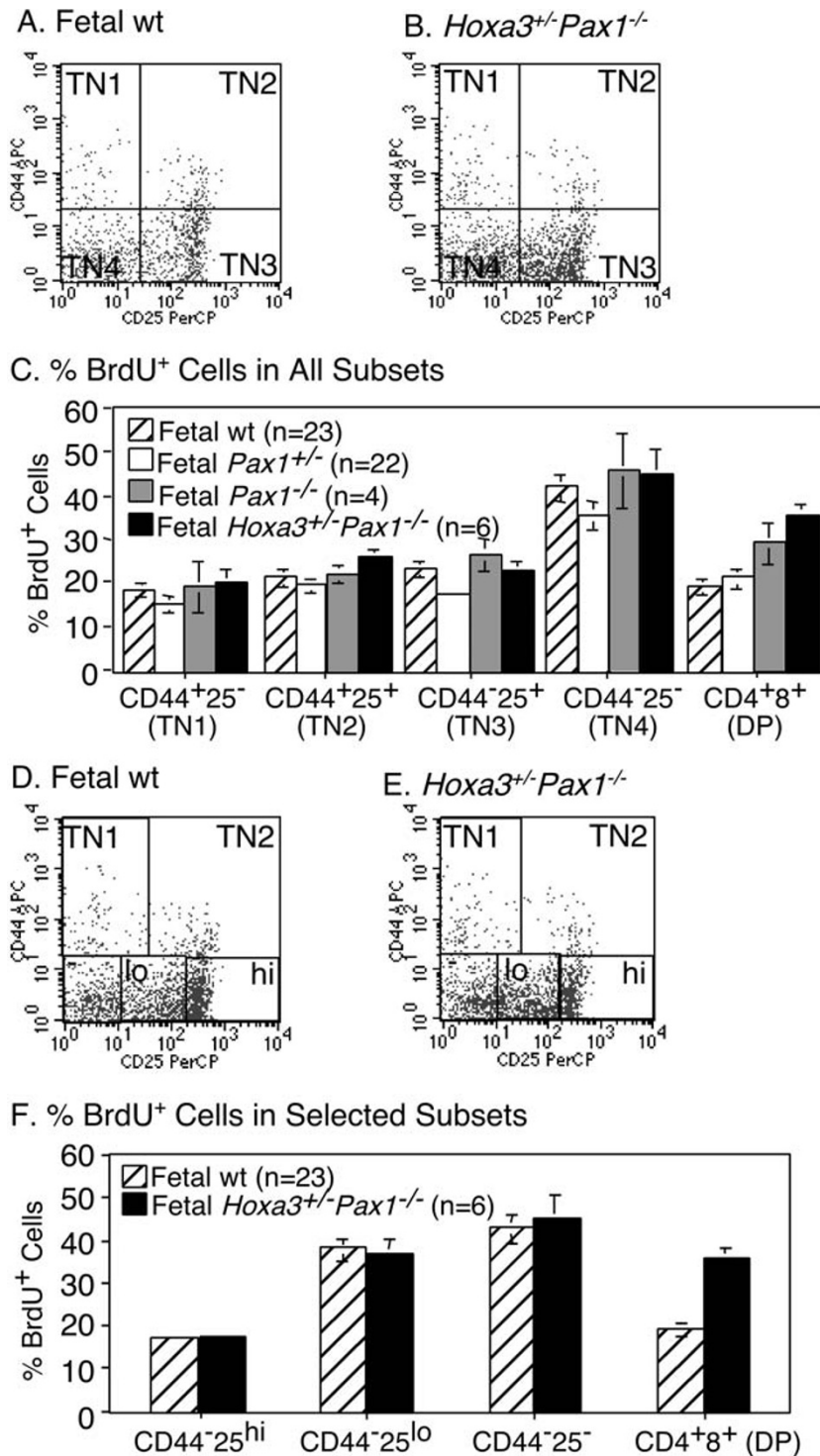


Figure 2
Proliferation of E17.5 *Hoxa3-Pax1* compound mutant thymocytes compared to wild type. Panels A-B show CD44 vs. CD25 dot plots of the gated CD3⁺4⁺8⁻ population from E17.5 wild type (A) and *Hoxa3*^{+/-}*Pax1*^{-/-} (B) thymus. Percentages are shown in each quadrant. C. Percentages of BrdU⁺ cells (Mean ± SEM) in each of 4 subsets of TN cells defined by CD44 and CD25, and in DP cells. D, E. CD44 vs. CD25 dot plots of the gated CD3⁺4⁺8⁻ population divided into five subsets as defined by [12]. F. Percentages of BrdU⁺ cells (Mean ± SEM) in each of 3 CD44⁻ TN subsets defined in panels D and E. DP values shown for comparison are the same as in C.

10-fold decrease in DP cell numbers [29]. Using BrdU analysis, we compared the proliferation of *Hoxa3^{+/-}Pax1^{-/-}* fetal thymocytes during the TN to DP transition compared to littermate controls. Thymocyte proliferation levels for all genotypes (*Hoxa3^{+/-}, Pax1^{+/-}, Pax1^{-/-}, and Hoxa3^{+/-}Pax1^{-/-}*) were similar to wild type in all of the canonical TN1-TN4 stages (Fig. 2A,2B,2C; Table 1), consistent with the normal percentages and cell numbers for these subpopulations previously reported [29]. Recent papers by Penit, et al. have provided data from cell cycle analysis suggesting that a discrete CD44⁺25^{lo} population with a specific proliferation profile can be identified, such that the transition from TN3 to DP can be further subdivided into CD25^{hi}, CD25^{lo}, and CD25⁻ subsets [12,13]. Analysis of proliferation using these more narrowly defined subsets still showed values similar to wild type in the CD44⁺25^{lo} subset (Fig. 2D,2E,2F; Table 2). In contrast, the proliferation level of DP cells increased nearly 2-fold in compound mutants compared to wild type (Fig. 2C, 2F; Table 1). In spite of this increase in the percentage of proliferating cells, the actual number of BrdU⁺ DP cells was reduced 4-fold compared to wild type in the compound mutants ($15.03 \pm 8.73 \times 10^4$ vs. $61.94 \pm 32.12 \times 10^4$ in wild type; $p < 0.01$). The numbers of proliferating cells at the pre-DP stage were unchanged (not shown). Combined with our previous results, this result shows that both increased apoptosis and reduced proliferation contribute to the decrease in DP cells in the *Hoxa3,Pax1* compound mutants. Unfortunately, *Hoxa3^{+/-}Pax1^{-/-}* mice die at birth, preventing analysis of thymocyte proliferation in adult compound mutants.

Cytokine production is decreased in the *Hoxa3^{+/-}Pax1^{-/-}* fetal thymus

Cytokines produced by TECs are good candidates for mediating at least some of these changes in thymocyte proliferation and apoptosis in the *Hoxa3,Pax1* compound mutants. Using semi-quantitative RT-PCR, we analyzed the expression of two cytokines that are produced by TECs, IL-7 and SCF, in pooled, genotype-matched E14.5 thymi (Fig. 3). *Hoxa3^{+/-}Pax1^{-/-}* compound mutants had decreased transcription of both cytokines relative to all other genotypes, including *Pax1^{-/-}* single mutants. However, *Hoxa3^{+/-}Pax1^{-/-}* compound mutants were not completely missing either cytokine analyzed, and the magnitude of the decrease was variable between pools.

To test whether the changes in cytokine levels were responsible for the observed thymocyte development phenotypes, we attempted to rescue these phenotypes by supplementation in fetal thymic organ culture (FTOC). E15.5 embryonic thymic lobes from individual embryos were cultured separately in FTOC, with or without added IL-7 or SCF. Cultures from control genotypes (wild type, *Pax1^{+/-}, Hoxa3^{+/-}, Pax1^{-/-}*) behaved as expected [41–46],

with addition of IL-7 causing an increase in the percentage of DN cells and a corresponding decrease in DP cells, while adding SCF had no significant effect on thymocyte development (data not shown). However, to our surprise the two separated lobes from *Hoxa3^{+/-}Pax1^{-/-}* compound mutants did not always develop in predictable ways. In fact, separated lobes cultured under identical conditions in the absence of any added cytokines had widely differing results. Changes in thymocyte cell number and differentiation after culture were highly variable; for example, cell numbers could differ stochastically after culture by as much as 10-fold between two lobes from the same compound mutant embryo. This result was never obtained with separately cultured thymic lobes from any other genotype, which always developed identically when cultured without cytokines. This property of the compound mutant thymic lobes probably reflects phenotypic variability previously seen in our histological and gross morphological analysis of these mutants [29,34].

Discussion

In this study, we have examined thymocyte proliferation in the fetal and adult wild type thymus. Our results showed clear stage-specific differences in wild type thymocyte proliferation depending on the developmental age of the microenvironment (i.e. fetal vs. adult). We also analyzed two mutant strains that have either cell-autonomous (*Hoxa3^{+/-}Pax1^{-/-}*) or non-cell-autonomous (*Rag1^{-/-}*) TEC defects, with blocks at different stages of thymocyte differentiation. Abnormal TEC development in both of these mutant strains resulted in both increases and decreases in thymocyte proliferation in a stage-specific manner, rather than simply an overall reduced ability to promote proliferation.

There is considerable evidence that thymopoiesis differs in the fetal thymus during the initial creation of the microenvironment, as opposed to adult thymus function (reviewed in [47]). Differences have been reported in responses to some cytokines, in signaling at the TN-DP transition in fetal compared to adult thymus, and in proliferation of CD4⁺ SP thymocytes [36,48–50]. The proliferation profiles of both wild type and *Rag1^{-/-}* fetal thymocytes support the hypothesis that fetal thymocyte maturation is fundamentally different from thymocyte development in the adult thymus. The TN1 and DP stages both show increased proliferation in wild-type fetal thymocytes compared to adults. This difference at the TN1 stage is not due to the presence of more B or NK cells in the adult thymus. Although it is formally possible that some other cell type may still be included in our TN1 population, this analysis strongly suggests that TN1 cells have different proliferation profiles in fetal and adult thymus.

Table 2: Percentages (Mean \pm SD) of BrdU+ cells in selected subsets of wild-type and compound mutant fetal thymocytes

Genotype	Total Cell #($\times 10^5$)	No.	CD44 ⁻ CD25 ^{hi}	CD44 ⁻ CD25 ^{lo}	CD44 ⁻ CD25 ⁻	CD4 ⁺ CD8 ⁺ CD3 ⁺
Wild type	42.72 \pm 3.88	23	16.91 \pm 5.77	37.65 \pm 12.77	42.45 \pm 15.73	19.05 \pm 8.12
<i>Hoxa3</i> ^{+/-} <i>Pax1</i> ^{-/-}	7.96 \pm 1.12	6	16.66 \pm 2.10	36.99 \pm 9.067	44.49 \pm 14.90	35.37 \pm 6.41**

**P < 0.01, significant difference with wild type.

It has been well documented that thymocytes in adult *Rag1* or *Rag2* mutants have dramatically reduced proliferation compared to wild type, particularly at the TN3 stage [37–40], and Penit et al. reported 2–10 fold reductions at every stage of thymocyte development in *Rag*^{-/-} adults [13]. Recent work by Petrie, et al. has shown that this reduced proliferation is not intrinsic to *Rag* mutant thymocytes, but rather is secondary to thymic micro-environment defects in *Rag* mutants [37]. Our results show that at fetal stages, *Rag* mutant thymocytes have a proliferation profile that is much more similar to wild type thymocytes at the same stage, with less than a 2-fold difference in the percentage of proliferating cells at any stage. For example, TN3 stage thymocytes in the *Rag*^{-/-} adult thymus have proliferation levels 3-fold below wild type thymus [13,37], while in the *Rag*^{-/-} fetal thymus TN3 cells are only 25–30% lower than in wild type fetal thymus. Proliferation is not changed at all stages, and is actually increased relative to wild type at the TN2 stage. This increase at the TN2 stage may be due to the failure of TN2 cells to initiate TCR β gene rearrangement at this stage, which is normally associated with decreased proliferation. *Rag*^{-/-} mutants have a profound thymocyte development block at the TN3 to TN4/preDP transition [51]. During normal wild type fetal thymus development, significant DP cell development does not occur until E16.5 in most genetic backgrounds. As a result, at E17.5 the thymic microenvironment in *Rag* mutants has not been exposed to abnormal thymocyte development for very long, and has not yet developed the full phenotype seen in the steady state adult *Rag* mutant thymus [35].

In the *Hoxa3*,*Pax1* compound mutants, cell-autonomous TEC defects cause a partial block at the DN-DP transition, so thymocytes do progress to DP and SP stages, albeit at reduced frequencies [29]. TEC defects are seen as early as E12.5 in these mutants, from the earliest stages of organogenesis [34]. Therefore, the TEC defects in these mice would be expected to have different effects on thymocyte proliferation than in *Rag* mutants. In this study, we found stage-specific changes in thymocyte proliferation at the DP stage. Our previous analysis reported decreased DP cell number and increased apoptosis in both the pre-DP

and DP stages in these mutants [29]. DP cell numbers in both this and our previous study were decreased 10-fold in the *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutants. At first glance, the increase in the percentage of proliferating cells at the DP stage appears contradictory to this phenotype. However, this increase actually corresponds to decreased numbers of proliferating DP cells compared to wild type. In contrast, pre-DP cells did not have a change in the number or percentage of proliferating cells, suggesting that the signals promoting proliferation of these cells is still present, and that proliferation of DP and pre-DP cells may be regulated independently. Pre-DP cells do have a three-fold increase in apoptosis [29], suggesting that separate signals regulate proliferation and survival of these cells. Combined with our previous data, this result suggests that the decrease in DP cell numbers seen in the *Hoxa3*-*Pax1* compound mutants is both due to an increase in the number of pre-DP and DP cells undergoing apoptosis as well as a decrease in the number of proliferating DP.

Insufficiencies of multiple cytokines might be involved in the changes in proliferation and cell death at the CD44⁻25⁻ and DP stages in *Hoxa3*-*Pax1* compound mutants. SCF and IL-7 are first expressed as early as E12.5 in the developing thymus, well before significant thymocyte maturation takes place [52,53]. Although the number of TECs is reduced by about 2-fold in *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutants, the number of thymocytes is reduced by a slightly greater amount, around 3-fold [29]. Therefore, the reductions seen in cytokine message levels in the *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutants are not secondary to decreased TEC numbers. In fact, these mutants should have a relative increase in the proportion of TECs, making the reductions seen in cytokine levels in the mutant thymi even more striking. The reduced expression of these cytokines in *Hoxa3*^{+/-}*Pax1*^{-/-} compound mutants could be an indirect effect of defective TEC differentiation, or could be direct downstream targets since both *Hoxa3* and *Pax1* are expressed in fetal thymus [54,55]. Comparison of the *Hoxa3*^{+/-}*Pax1*^{-/-} phenotype with those of cytokine mutants do not reveal many similarities [30,46,56–61], probably because of the simultaneous reductions in multiple cytokines in the *Hoxa3*^{+/-}*Pax1*^{-/-} mutants. For example, SCF

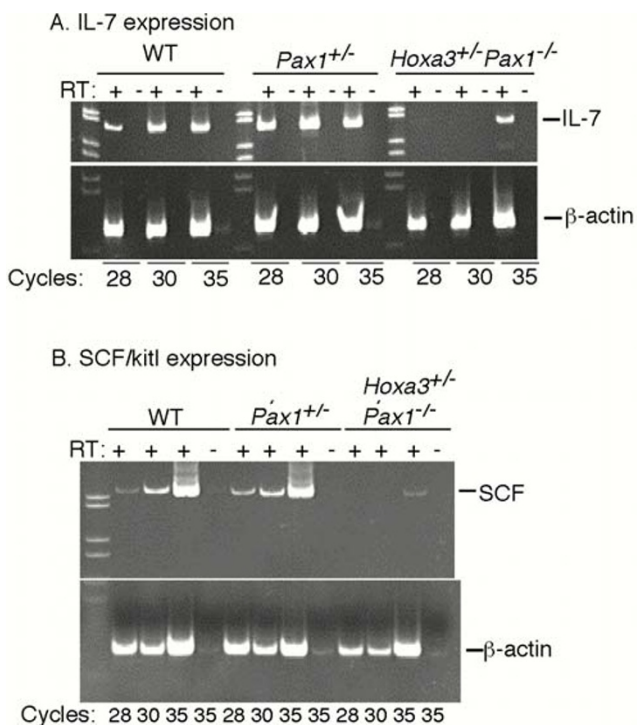


Figure 3
Expression of cytokines in fetal thymi of *Hoxa3*^{+/-}*Pax1*^{-/-} mice and controls measured by semi-quantitative RT-PCR Each lane contains pooled cDNA from 5–10 E14.5 embryonic thymi of the same genotype. Representative results from one of three experiments for each cytokine are shown. β -actin was used as a control. Panel A, expression of IL-7; Panel B, expression of SCF.

and IL-7 have been shown to have synergistic functions [62,63]. Therefore, changes in multiple cytokines in *Hoxa3*^{+/-}*Pax1*^{-/-} mutants would be expected to have complex effects on thymocyte development that could be difficult to predict.

Interpretation of the observed phenotype is further complicated by the fact that none of the cytokines tested are completely absent. A striking example is the complete absence of $\gamma\delta$ TCR⁺ thymocyte development in IL-7 mutants, in contrast to normal $\gamma\delta$ TCR⁺ cell development in *Hoxa3*^{+/-}*Pax1*^{-/-} mutants [29], which have dramatically reduced IL-7 levels. This result suggests that although $\gamma\delta$ TCR⁺ cell development requires IL-7, low levels are sufficient for this function. Therefore, analysis of mutants with low levels of cytokines may provide a system for uncovering both dosage-dependent requirements and later functions for these factors in promoting thymocyte development. It is also possible that some of the differences seen between *Hoxa3*^{+/-}*Pax1*^{-/-} and cytokine mutant phenotypes reflect different effects of cytokines at fetal and adult stages. Stud-

ies of IL-7R mutants showed relatively normal thymocyte maturation at fetal stages, albeit with reduced thymocyte numbers, in contrast to a severe block in thymocyte differentiation in adult thymus [49]. Analysis of cytokine mutants during fetal stages could therefore reveal a more complete picture of the role of these cytokines during fetal thymus development.

Conclusions

Our results show that fetal and adult wild type thymocytes have stage-specific differences in proliferation, with both TN1 and DP cells exhibiting higher levels of proliferation in fetal thymus. This fetal wild-type profile was very similar to that seen for fetal *Rag* mutant thymocytes, in contrast to adult *Rag*^{-/-} thymus. Therefore, the changes in proliferation seen in the fetal *Rag* mutant thymocytes reflect the earliest microenvironment changes occurring as a result of defective TEC-thymocyte interactions in the *Rag*^{-/-} thymus.

The current study further supports a role for TECs in regulating both thymocyte proliferation and survival during the DN-DP transition, and suggests that proliferation and apoptosis of TN4 and DP cells are regulated independently. Our results provide direct genetic evidence for these functions of TECs, by showing that these processes are defective in mutants with a primary TEC differentiation defect (the *Hoxa3*,*Pax1* compound mutants). Taken together, our results further define the role TECs play at this complex stage of thymocyte development.

Methods

Mice and genotyping

Hoxa3^{+/-}*Pax1*^{+/-} mice were mated to *Pax1*^{+/-} or *Pax1*^{-/-} mice and genotyped for both *Hoxa3* and *Pax1* by PCR as described previously [29]. Comparisons were made between littermates from the same colony. *Rag1*^{-/-} mice (Jackson Labs) were maintained as a homozygous mutant colony. All mice were maintained as congenic on a C57BL/6 genetic background. Noon of the day of vaginal plug was considered E0.5. All animal use procedures were approved by the Medical College of Georgia Committee on Animal Use for Research and Education (CAURE).

Flow cytometry

Fetal thymocyte in vivo BrdU incorporation was done by intraperitoneal injection of BrdU (5-Bromo-2'-Deoxyuridine; Sigma) on the morning of E17.5, two doses per mouse and 1 mg per dose with a 0.5 hour pause, as previously described [13]. Two hours after the first injection, fetal thymi were isolated by dissection and single-cell suspensions prepared by passing the thymocytes through cell strainers. Adult thymocyte in vivo BrdU incorporation was done by the same injection protocol using 6–8 week old C57BL/6 adult mice.

Four-color flow cytometry was performed using the following monoclonal antibodies (Pharmingen): biotin anti-mouse CD25 (clone: 7D4) visualized with Per-CP conjugated streptavidin; APC anti-mouse CD44 (clone: IM7); and three phycoerythrin (PE) conjugated antibodies: anti-mouse CD4 (clone: RM4-5), anti-mouse CD8a (clone: 53-6.7), and anti-mouse CD3 ϵ (clone: 145-2C11). In some experiments, PE conjugated antibodies against B220 (clone: RA3-6B2) and NK1.1 (clone: PK136) were also included. After cell surface staining, the cells were fixed with 2% paraformaldehyde (PFA)/PBS at 4°C overnight. The cells were then washed, permeabilized with 0.1% Triton X-100/ 0.1% Na-Citrate (pH7.2) on ice for 2 min, and then treated with 50 units of DNaseI at 37°C for 10 min. Cells were then stained with FITC anti-BrdU antibody (Pharmingen). Data were collected by selectively gating either CD3⁺4⁻8⁻ or CD3⁺4⁺8⁺ populations (Becton Dickinson dual laser FACSCalibur), and analyses were performed with Cellquest software. Significant differences were determined by the unpaired Student's t-test or ANOVA analysis.

Semi-quantitative RT-PCR

E14.5 fetal thymic lobes were collected and stored in the TRIzol (GIBCO BRL) at -70°C until RNA isolation. Pools of 5-to10 thymi from embryos of the same genotype were homogenized using Micro Pellet Pestles (Nalge Nunc International) in 1.5 ml-ependorf tubes. RNA was isolated and the genomic DNA was depleted by using DNaseI (Amplification Grade, GIBCO BRL) following the manufacturer's protocols. Reverse transcription of 1 μ g of total RNA to cDNA was done using SuperScript II following the manufacturer's protocol (GIBCO BRL). Equal amounts of cDNA from different genotypes were added to a final 20 μ l PCR reaction mixture using the QIAGEN Taq PCR kits. PCR conditions were: 94°C 30 sec, 50°C 1 min, 72°C 1 min for IL-7, SCF and their internal control β -actin; or 94°C 30 sec, 60°C 1 min, 72°C 1 min for TGF β 1 and its internal control β -actin. PCR products were visualized by 5% acrylamide gel electrophoresis and ethidium bromide staining. Band densities were measured and analyzed using the Molecular Analyst software package (Bio-Rad Laboratories, Version 1.4.1). Primer sequences were: IL-7 [65]: IL-7 (5') 5'ACT ACA CCC ACC TCC CGC A3'; IL-7 (3') 5'TCT CAG TAG TCT CTT TAG G3'; SCF [65]: SCF (5') 5'TCT TCA ACT GCT CCT ATT T3'; SCF (3') 5'ACT GCT ACT GCT GTC ATT C3'; TGF β 1 [66]: TGF β 1(5') 5'GCG GAC TAC TAT GCT AAA GAG G3'; TGF β 1(3') 5'GTT GTG TTG GTT GTA GAG GGC A3'; β -actin [67]: β -actin (5') 5'GGG TCA GAA GGA CTC CTA TG3'; β -actin (3') 5'GTA ACA ATG CCA TGT TCA AT3'.

Fetal thymic organ culture (FTOC)

Thymic lobes were dissected from E15.5 embryos. The two lobes from each embryo cultured separately in high

oxygen submersion culture as previously described [41]. Fetal thymic organ cultures were supplemented with 100 u/ml of IL-7 or 20 ng/ml of SCF (Pepro Tech Inc.) as previously described [41]. For each embryo, one lobe was treated while the other was cultured in parallel without supplemental cytokine as an internal control. In control experiments, lobes were cultured separately under identical conditions without cytokine. After five days in culture, the cells were stained by PE-anti-mouse CD4 and FITC-anti-mouse CD8 and analyzed by flow cytometry as above.

Authors' contributions

DMS conceived of the study and performed all of the experiments described. NRM contributed to the design of the study and data analysis, and drafted the manuscript.

All authors read and approved the final manuscript.

Acknowledgements

We thank Sammy Navarre and Lili Cheng for technical contributions. This work was supported by National Institutes of Health Grant# R01-HD35920 to NRM.

References

- Boyd RL, Tucek CL, Godfrey DI, Izon DJ, Wilson TJ, Davidson NJ, Bean AG, Ladyman HM, Ritter MA, Hugo P: **The thymic microenvironment.** *Immunol Today* 1993, **14**:445-459
- Anderson G, Harman BC, Hare KJ, Jenkinson EJ: **Microenvironmental regulation of T cell development in the thymus.** *Semin Immunol* 2000, **12**:457-464
- Ritter MA, Boyd RL: **Development in the thymus: it takes two to tango.** *Immunol Today* 1993, **14**:462-469
- van Ewijk W, Hollander G, Terhorst C, Wang B: **Stepwise development of thymic microenvironments in vivo is regulated by thymocyte subsets.** *Development* 2000, **127**:1583-1591
- van Ewijk W, Kawamoto H, Germeaad WT, Katsura Y: **Developing thymocytes organize thymic microenvironments.** *Curr Top Microbiol Immunol* 2000, **251**:125-132
- Anderson G, Moore NC, Owen JJ, Jenkinson EJ: **Cellular interactions in thymocyte development.** *Ann Rev Immunol* 1996, **14**:73-99
- Godfrey DI, Zlotnik A: **Control points in early T-cell development.** *Immunol Today* 1993, **14**:547-553
- Godfrey DI, Kennedy J, Suda T, Zlotnik A: **A developmental pathway involving four phenotypically and functionally distinct subsets of CD3-CD4-CD8- triple-negative adult mouse thymocytes defined by CD44 and CD25 expression.** *J Immunol* 1993, **150**:4244-4252
- von Boehmer H, Aifantis I, Feinberg J, Lechner O, Saint-Ruf C, Walter U, Buer J, Azogui O: **Pleiotropic changes controlled by the pre-T-cell receptor.** *Curr Opin Immunol* 1999, **11**:135-142
- Rothenberg EV: **The development of functionally responsive T cells.** *Advances in Immunology* 1992, **51**:85-214
- Jameson SC, Hogquist KA, Bevan MJ: **Positive selection of thymocytes.** *Annu Rev Immunol* 1995, **13**:93-126
- Vasseur F, Le Campion A, Penit C: **Scheduled kinetics of cell proliferation and phenotypic changes during immature thymocyte generation.** *Eur J Immunol* 2001, **31**:3038-3047
- Penit C, Lucas B, Vasseur F: **Cell expansion and growth arrest phases during the transition from precursor (CD4⁺8⁻) to immature (CD4⁺) J Immunol 1995, **154**:5103-5113**
- Penit C, Vasseur F: **Cell proliferation and differentiation in the fetal and early postnatal mouse thymus.** *J Immunol* 1989, **142**:3369-3377
- Hare KJ, Wilkinson RW, Jenkinson EJ, Anderson G: **Identification of a developmentally regulated phase of postselection expansion**

- sion driven by thymic epithelium. *J Immunol* 1998, **160**:3666-3672
16. Jotereau FV, Heuze F, Salamon-Vie V, Gascon H: **Cell kinetics in the fetal mouse thymus: precursor cell input, proliferation, and emigration.** *J Immunol* 1987, **138**:1026-1030
 17. Falk I, Biro J, Kohler H, Eichmann K: **Proliferation kinetics associated with T cell receptor- β chain selection of fetal murine thymocytes.** *J Exp Med* 1996, **184**:2327-2339
 18. Parkin IG, Owen JJ, Jenkinson EJ: **Proliferation of thymocytes in relation to T-cell receptor beta-chain expression.** *Immunology* 1988, **64**:97-99
 19. Anderson G, Jenkinson EJ, Moore NC, Owen JJ: **MHC class II-positive epithelium and mesenchyme cells are both required for T-cell development in the thymus.** *Nature* 1993, **362**:70-73
 20. Markowitz JS, Auchincloss H Jr, Grusby MJ, Glimcher LH: **Class II-positive hematopoietic cells cannot mediate positive selection of CD4⁺ T lymphocytes in class II-deficient mice.** *Proc Natl Acad Sci USA* 1993, **90**:2779-2783
 21. Laufer TM, DeKoning J, Markowitz JS, Lo D, Glimcher LH: **Unopposed positive selection and autoreactivity in mice expressing class II MHC only on thymic cortex.** *Nature* 1996, **383**:81-85
 22. Jenkinson EJ, Anderson G, Moore NC, Smith CA, Owen JJ: **Positive selection by purified MHC class II⁺ thymic epithelial cells in vitro: costimulatory signals mediated by B7 are not involved.** *Dev Immunol* 1994, **3**:265-271
 23. Di Santo JP, Rodewald H-R: **In vivo roles of receptor tyrosine kinases and cytokine receptors in early thymocyte development.** *Curr Opin Immunol* 1998, **10**:196-207
 24. Malek TR, Porter BO, He Y-W: **Multiple γ c-dependent cytokines regulate T-cell development.** *Immunol Today* 1999, **20**:71-76
 25. MacDonald H, Budd R, Howe R: **A CD3⁻ subset of CD4⁺ thymocytes: a rapidly cycling intermediate in the generation of CD4⁺8⁺ cells.** *Eur J Immunol* 1988, **18**:519-523
 26. Petrie HT, Pearse M, Scollay R, Shortman K: **Development of immature thymocytes: initiation of CD3, CD4, and CD8 acquisition parallels down-regulation of the interleukin 2 receptor alpha chain.** *Eur J Immunol* 1990, **20**:2813-2815
 27. Nikolic-Zugic J, Moore MV, Bevan MJ: **Characterization of the subset of immature thymocytes which can undergo rapid in vitro differentiation.** *Eur J Immunol* 1989, **19**:649-653
 28. Petrie HT, Hugo P, Scollay R, Shortman K: **Lineage relationships and developmental kinetics of immature thymocytes: CD3, CD4, and CD8 acquisition in vivo and in vitro.** *J Exp Med* 1990, **172**:1583-1588
 29. Su DM, Manley NR: **Hoxa3 and pax1 transcription factors regulate the ability of fetal thymic epithelial cells to promote thymocyte development.** *J Immunol* 2000, **164**:5753-5760
 30. Takahama Y, Letterio JJ, Suzuki H, Farr AG, Singer A: **Early progression of thymocytes along the CD4/CD8 developmental pathway is regulated by a subset of thymic epithelial cells expressing Transforming Growth Factor β .** *J Exp Med* 1994, **179**:1495-1506
 31. Manley NR, Capecchi MR: **Hox group 3 paralogs regulate the development and migration of the thymus, thyroid, and parathyroid glands.** *Dev Biol* 1998, **195**:1-15
 32. Manley NR, Capecchi MR: **The role of Hoxa-3 in mouse thymus and thyroid development.** *Development* 1995, **121**:1989-2003
 33. Wallin J, Eibel H, Neubuser A, Wilting J, Koseki H, Balling R: **Pax1 is expressed during development of the thymus epithelium and is required for normal T-cell maturation.** *Development* 1996, **122**:23-30
 34. Su D, Ellis S, Napier A, Lee K, Manley NR: **Hoxa3 and pax1 regulate epithelial cell death and proliferation during thymus and parathyroid organogenesis.** *Dev Biol* 2001, **236**:316-329
 35. Naquet P, Naspetti M, Boyd R: **Development, organization and function of the thymic medulla in normal, immunodeficient or autoimmune mice.** *Semin Immunol* 1999, **11**:47-55
 36. Ceredig R: **Intrathymic proliferation of perinatal mouse alpha beta and gamma delta T cell receptor-expressing mature T cells.** *Int Immunol* 1990, **2**:859-867
 37. Petrie HT, Tourigny M, Burtrum DB, Livak F: **Precursor thymocyte proliferation and differentiation are controlled by signals unrelated to the pre-TCR.** *J Immunol* 2000, **165**:3094-3098
 38. Tourigny MR, Mazel S, Burtrum DB, Petrie HT: **T cell receptor (TCR)- β gene recombination: dissociation from cell cycle regulation and developmental progression during T cell ontogeny.** *Journal of Experimental Medicine* 1997, **185**:1549-1556
 39. Hoffman ES, Passoni L, Crompton T, Leu TM, Schatz DG, Koff A, Owen MJ, Hayday AC: **Productive T-cell receptor beta-chain gene rearrangement: coincident regulation of cell cycle and clonality during development in vivo.** *Genes Dev* 1996, **10**:948-962
 40. Diamond RA, Ward SB, Owada-Makabe K, Wang H, Rothenberg EV: **Different developmental arrest points in RAG-2^{-/-} and SCID thymocytes on two genetic backgrounds: developmental choices and cell death mechanisms before TCR gene rearrangement.** *J Immunol* 1997, **158**:4052-4064
 41. Su D-M, Wang J, Lin Q, Cooper MD, Watanabe T: **Interferons α/β inhibit IL-7-induced proliferation of CD4⁺CD8⁻CD3⁻CD44⁺CD25⁺ thymocytes, but do not inhibit that of CD4⁺CD8⁻CD3⁻CD44⁻CD25⁻ thymocytes.** *Immunology* 1997, **90**:543-549
 42. Moore TA, Zlotnik A: **T-cell lineage commitment and cytokine responses of thymic progenitors.** *Blood* 1995, **86**:1850-1860
 43. Murray R, Suda T, Wrighton N, Lee F, Zlotnik A: **IL-7 is a growth and maintenance factor for mature and immature thymocyte subsets.** *Int Immunol* 1989, **1**:526-531
 44. Watson J, Morrissy P, Namen A, Conlon P, Widmer M: **Effect of IL-7 on the growth of fetal thymocytes in culture.** *J Immunol* 1989, **143**:1215-1222
 45. Plum J, De Smedt M, Leclercq G: **Exogenous IL-7 promotes the growth of CD3⁻CD4⁺CD8⁻CD44⁺CD25⁺ precursor cells and blocks the differentiation pathway of TCR-ab cells in fetal thymic organ culture.** *J Immunol* 1993, **150**:2706-2716
 46. Plum J, De Smedt M, Leclercq G, Vandekerckhove B: **Influence of TFG- β on murine thymocyte development in fetal thymus organ culture.** *J Immunol* 1995, **154**:5789-5798
 47. Manley N: **Thymus organogenesis and molecular mechanisms of thymic epithelial cell differentiation.** *Seminars in Immunology* 2000, **12**:421-428
 48. Ceredig R: **Proliferation in vitro and interleukin production by 14 day fetal and adult Lyt-2/L3T4- mouse thymocytes.** *J Immunol* 1986, **137**:2260-2267
 49. Crompton T, Outram SV, Buckland J, Owen MJ: **Distinct roles of the interleukin-7 receptor alpha chain in fetal and adult thymocyte development revealed by analysis of interleukin-7 receptor alpha-deficient mice.** *Eur J Immunol* 1998, **28**:1859-1866
 50. Molina TJ, Perrot JY, Penninger J, Ramos A, Audouin J, Briand P, Mak TW, Diebold J: **Differential requirement for p56lck in fetal and adult thymopoiesis.** *J Immunol* 1998, **160**:3828-3834
 51. Mombaerts P, Iacomini J, Johnson RS, Herrup K, Tonegawa S, Papaioannou VE: **RAG -I-deficient mice have no mature B and T lymphocytes.** *Cell* 1992, **68**:869-877
 52. Moore NC, Anderson G, Smith CA, Owen JJ, Jenkinson EJ: **Analysis of cytokine gene expression in subpopulations of freshly isolated thymocytes and thymic stromal cells using semiquantitative polymerase chain reaction.** *Eur J Immunol* 1993, **23**:922-927
 53. Wiles MV, Ruiz P, Imhof BA: **Interleukin-7 expression during mouse thymus development.** *Eur J Immunol* 1992, **22**:1037-1042
 54. Wallin J, Eibel H, Neubuser A, Wilting J, Koseki H, Balling R: **Pax1 is expressed during development of the thymus epithelium and is required for normal T-cell maturation.** *Development* 1996, **122**:23-30
 55. Su D, Ellis S, Napier A, Lee K, Manley NR: **Hoxa3 and pax1 regulate epithelial cell death and proliferation during thymus and parathyroid organogenesis.** *Dev Biol* 2001, **236**:316-329
 56. Rodewald H-R, Kretzschmar K, Swat W, Takeda S: **Intrathymically expressed c-kit ligand (stem cell factor) is a major factor driving expansion of very immature thymocytes in vivo.** *Immunity* 1995, **3**:313-319
 57. Porter BO, Scibelli P, Malek TR: **Control of T cell development in vivo by subdomains within the IL-7 receptor alpha-chain cytoplasmic tail.** *J Immunol* 2001, **166**:262-269
 58. Malek TR, Porter BO, He YW: **Multiple gamma c-dependent cytokines regulate T-cell development.** *Immunol Today* 1999, **20**:71-76
 59. Bhatia SK, Tygrett LT, Grabstein KH, Waldschmidt TJ: **The effect of in vivo IL-7 deprivation on T cell maturation.** *J Exp Med* 1995, **181**:1399-1409

60. Peschon JJ, Morrissey PJ, Grabstein KH, Ramsdell FJ, Maraskovsky E, Gliniak BC, Park LS, Ziegler SF, Williams DE, Ware CB, Meyer JD, Davison BL: **Early lymphocyte expansion is severely impaired in interleukin 7 receptor-deficient mice.** *Journal of Experimental Medicine* 1994, **180**:1955-1960
61. Maki K, Sunaga S, Komagata Y, Kodaira Y, Mabuchi A, Karasuyama H, Yokomuro K, Miyazaki J-I, Ikuta K: **Interleukin 7 receptor-deficient mice lack $\gamma\delta$ T cells.** *Proc Natl Acad Sci USA* 1996, **93**:7172-7177
62. Morrissey PJ, McKenna H, Widmer MB, Braddy S, Voice R, Charrier K, Williams DE, Watson JD: **Steel factor (c-kit ligand) stimulates the in vitro growth of immature CD3⁺CD4⁺CD8⁻ thymocytes: synergy with IL-7.** *Cell Immunol* 1994, **157**:118-131
63. Rodewald H-R, Ogawa M, Haller C, Waskow C, DiSanto JP: **Pro-thymocyte expansion by c-kit and the common cytokine receptor γ chain is essential for repertoire formation.** *Immunity* 1997, **6**:265-272
64. Chantry D, Turner M, Feldmann M: **Interleukin 7 (murine pre-B cell growth factor/lymphopoietin 1) stimulates thymocyte growth: regulation by transforming growth factor beta.** *Eur J Immunol* 1989, **19**:783-786
65. Moore NC, Anderson G, Smith CA, Owen JJ, Jenkinson EJ: **Analysis of cytokine gene expression in subpopulations of freshly isolated thymocytes and thymic stromal cells using semiquantitative polymerase chain reaction.** *Eur J Immunol* 1993, **23**:922-927
66. Shull MM, Ormsby I, Kier AB, Pawlowski S, Diebold RJ, Yin M, Allen R, Sidman C, Proetzel G, Calvin D, et al: **Targeted disruption of the mouse transforming growth factor-beta 1 gene results in multifocal inflammatory disease.** *Nature* 1992, **359**:693-699
67. Mansour SL, Goddard JM, Capecchi MR: **Mice homozygous for a targeted disruption of the proto-oncogene int-2 have developmental defects in the tail and inner ear.** *Development* 1993, **117**:13-28

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMedcentral will be the most significant development for disseminating the results of biomedical research in our lifetime."

Paul Nurse, Director-General, Imperial Cancer Research Fund

Publish with **BMC** and your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours - you keep the copyright



BioMedcentral.com

Submit your manuscript here:

<http://www.biomedcentral.com/manuscript/>

editorial@biomedcentral.com